

Impact of different phytochemical classes and *Ayurvedic* plants in battle against cancer

Bhagyashri Vaghora, Vinay Shukla

Department of Pharmaceutical Chemistry, Gujarat Ayurved University, Jamnagar 361 006, India

Cancer is believed the most dangerous diseases throughout the world. *Ayurvedic* texts have mentioned this disease as *arbuda* and plants are also listed for treatment. Coordination between the *ayurvedic* and modern framework can be helpful for future studies regarding the drug discovery from these plants. In recent years, many plants have been screened for anticancer activity and numerous potential dynamic constituents have also been confined from them. This survey highlights the fundamental phytochemical classes taking part effectively against malignancy. The present paper discusses the current status of researches in the field of cancer with respect to *Ayurvedic* plants and focuses on the phytochemical classes potentially active as anticancer.

Introduction

Cancer remains one of the leading causes of morbidity and mortality globally. Amongst the noncommunicable diseases, cancer is the second leading cause of death, after cardiovascular disease. Cancer is responsible for one in eight deaths worldwide—more than AIDS, tuberculosis, and malaria together. Globally, the number of cancer death is projected to increase from 7.1 million in 2002 to 11.5 million in 2030.¹ Cancer is the uncontrolled growth of abnormal cells in the body. Cancerous cells are also called malignant cells. Although cancer can develop in virtually any of the body's tissues, and each type of cancer has its unique features, the basic processes that produce cancer are quite similar in all forms of the disease.²

Ayurveda, the oldest Indian indigenous medicine system of plant drug is known from very early times for preventing or suppressing various tumors using these natural drugs. And nowadays scientists are keener to researches on complementary and alternative medicine for the management of cancer. In *Ayurvedic* concept, according to '*Charaka*' and '*Sushruta Samhitas*' cancer is described as inflammatory or non-inflammatory swelling and mentioned either as '*Granthi*' (Minor neoplasm) or '*Arbuda*' (Major neoplasm).³ *Ayurvedic* literature defines three body-control systems, viz., the nervous system (*vata* or air), the venous system (*Pitta* or fire) and the arterial system (*Kapha* or water) which mutually coordinate to perform the normal function of the body. In benign neoplasm (*Vataja*, *Pittaja* or *Kaphaja*) one or two of the three bodily systems are out of control and is not too harmful because the body is still trying to coordinate among these systems. Malignant tumors (*Tridoshaja*) are very harmful because all the three major bodily system lose mutual coordination and thus cannot prevent tissue damage, resulting in a deadly morbid condition.⁴

Plants are natural sources of phytochemicals which are helpful to fight against diseases. The major phytochemicals to produce a significant pharmacological actions are tannins, alkaloids, flavonoids and flavones, steroids and sterols, phytohormones, terpenoids, natural pigments such as carotenoids, alpha and beta carotenes, saponins and sapogenins, cardiac glycosides etc. These phytochemicals play crucial role in fighting against the one of the dangerous disease cancer.

The anticancer activities of number of plants are still being actively researched and some have shown promising results. Here some most frequently examined plants are reviewed regarding their effect on cancer. Table 1 illustrates list of plants which have proven anticancer activity.

Table 1. Anticancer activity of plants

Sr. No.	Ayurvedic Name	Scientific Name	Family	Cancer cell line/Anticancer activity	References
1.	<i>Gunja</i>	<i>Abrus precatorius</i>	Leguminosae	Yoshida sarcoma (rats) Fibro sarcoma(mice), Ascites tumors cells	5
				Human breast cancer cell line MDA-MB-231	6
2.	<i>Bilva</i>	<i>Aegle marmelos</i>	Rutaceae	Ehrlich ascites carcinoma	7
				Breast cancer cell lines MCF7, MDA-MB-231	8
				Human cancer cell lines of lung (A549), colon (CoLo05), ovary (IGROV1), prostrate (PC3), leukaemia (THP1) and breast(MCF7) cancer	9
3.	<i>Ajmoda</i>	<i>Apium graveolens</i>	Umbelliferae	Dalton's lymphoma ascites (DLA) cells, L929 cell lines (Lung fibroblast)	10
4.	<i>Rasna</i>	<i>Alpinia galangal</i>	Zingiberaceae	A549(lung adenocarcinoma cells), CRL2522(normal fibroblast cells),MCF-12A(normal epithelial cells derived from breast), CRL2321(mammary carcinoma cells) and CRL2335(mammary carcinoma cells)	11
5.	<i>Shirish</i>	<i>Albizzia lebeck</i>	Mimosaceae	Sarcoma 180 (mice)	5
6.	<i>Palandu</i>	<i>Allium cepa</i>	Liliaceae	Human breast cancer MDA-MB-231 cells	12
7.	<i>Rasona</i>	<i>Allium sativum</i>	Liliaceae	Sarcoma (rat)	5
8.	<i>Kumari</i>	<i>Aloe vera</i>	Liliaceae	Yoshida AH-130 ascites hepatoma (pleural tumour) human neuroectodermal tumors	5
	<i>Saptaparna</i>	<i>Alstonia scholaris</i>	Apocyanaceae	HSI human sarcoma	5
				Benzo(a)pyrene induced forestomach carcinoma	13
				Skin carcinogenesis(Mice)	14
9.	<i>Surana</i>	<i>Amorphopallus campanulates</i>	Araceae	Colon carcinoma cell line HCT-15	15
10.	<i>Kalmegh</i>	<i>Andrographis paniculata</i>	Acanthaceae	Neuroblastoma (IMR-32) And Human Colon (HT-29) Cancer Cell Line	16
11.	<i>Lakshamanap hala</i>	<i>Annona muricata</i>	Annonaceae	Breast cancer, T37D cell line	17
				Capan-1 Cells	18
				Skin Papillomagenesis in Mice	19
				HeLa cervical cancer cell	20
				Cell lines of EACC, MDA and SKBR3	21
12.	<i>Shatavari</i>	<i>Asparagus racemosa</i>	Liliaceae	Human epidermoid carcinoma	5
				UOK146 renal cell carcinoma cell line	22
13.	<i>Neem</i>	<i>Azadirachta indica</i>	Meliaceae	Prostate cancer, buccal carcinogenesis, , breast cancer, gastric carcinogenesis and B12 melanoma	23

14.	<i>Brahmi</i>	<i>Bacopa monnieri</i>	Scrophulariaceae	Walker carcinosarcoma 256 Human breast cancer (MCF7 and MDAMB 231) cell lines	5 24
15.	<i>Amritavallari</i>	<i>Basella rubra</i>	Basellaceae	Colon carcinogenesis Human cervical carcinoma	25 26
16.	<i>Kanchnar</i>	<i>Bauhinia variegata</i>	Caesalpinaceae	Prostrate (DU-145), lungs (HOP-62), ovary (IGR-OV-1), breast (MCF-7), and leukemia (THP-1) cell lines	27
17.	<i>Daruharidra</i>	<i>Berberis aristata</i>	Berberidaceae	Human epidermal carcinoma of the nasopharynx, <i>N</i> -nitrosodiethylamine induced Carcinogenesis	5
18.	<i>Shallaki</i>	<i>Boswellia serrata</i>	Burseraceae	Human epidermal carcinoma of the nasopharynx, Leukemia and brain tumors	5
19.	<i>Arka</i>	<i>Calotropis gigantea</i>	Asclepiadaceae	Human epidermal carcinoma of the Nasopharynx1	5
20.	<i>Senna</i>	<i>Cassia angustifolia</i>	Caesalpinaceae	Colorectal cancer	28
21.	<i>Indravaruni</i>	<i>Citrullus colocynthis</i>	Cucurbitaceae	Hepatocarcinoma(Hep G2 cell line)	29
22.	<i>Nimbuka</i>	<i>Citrus medica</i>	Rutaceae	Human astrocytoma cancer cell line (line1321)	30
23.	<i>Kesar</i>	<i>Crocus sativus</i>	Iridaceae	Gastric cancer, Colorectal cancer, Hepatic cancer, Pancreatic cancer, Prostate cancer, Cervical, ovarian, and breast cancer, Skin cancer, Lung cancer, Leukemia	31
24.	<i>Haridra</i>	<i>Curcuma longa</i>	Zingiberaceae	Fibro sarcoma, Preclinical and clinical trials B164A5 murine melanoma cells Dalton Lymphoma ascites (DLA) tumor model in Swiss Albino mice	5 32 33
25.	<i>Musta</i>	<i>Cyperus rotundus</i>	Cyperaceae	L1210 cells murine lymphoblastic leukaemia K562 and L1210 cell lines K562 erythroleukemia cells	34 35 36
26.	<i>Dhatura</i>	<i>Datura metel</i>	Solanaceae	Human epidermal carcinoma of the Nasopharynx	5
27.	<i>Amalaki</i>	<i>Embllica officinalis</i>	Euphorbiaceae	Skin carcinogenesis Cervical and ovarian cancer cells, human liver cancer cell line HepG2 Human Ovarian Cancer Cell Proliferation	37 38 39
28.	<i>Paribhadra</i>	<i>Erythrina suberosa</i>	Leguminosae	Sarcoma 180	5
29.	<i>Dugdhika</i>	<i>Euphorbia hirta</i>	Euphorbiaceae	Freund virus leukemia	5
30.	<i>Udumbar</i>	<i>Ficus racemosa</i>	Moraceae	Renal carcinogenesis	40
31.	<i>Ashwatha</i>	<i>Ficus religiosa</i>	Moraceae	Human breast cancer cell line	41
32.	<i>Langli</i>	<i>Gloriosa superba</i>	Liliaceae	Human hepatoma cell lineHepG2 cells	42
33.	<i>Jimutaka</i>	<i>Luffa echinata</i>	Cucurbitaceae	colon cancer cells (HT-29)	5
34.	<i>Kampilak</i>	<i>Mallotus philippinensis</i>	Euphorbiaceae	A-549, COLO-205, DU-145, HEP-2, HeLa, IMR-32, KB,	43

				MCF-7, NCI-H23, OVCAR-5, SiHa, SK-N-MC, SW-620 AND ZR-75-1 cancer lines	
				HL-60 Leukemia cells	44
35.	<i>Amra</i>	<i>Mangifera indica</i>	Anacardiaceae	Adenocarcinoma cell lines	5
				Human breast cancer (MCF-7 and MDA-MB-231 cell lines)	45
				Human proximal tubule cell line	46
36.	<i>Karvellak</i>	<i>Momordica charantia</i>	Cucurbitaceae	A-549, COLO-205, MCF-7, NCI-H322, PC-3, THP-1 AND U-87MG	47
37.	<i>Shigru</i>	<i>Moringa oleifera</i>	Moringaceae	Human epidermoid lymphocytic leukemia Skin Papillomagenesis	5
				Cervical carcinoma, HeLa cell line	29
38.	<i>Karveer</i>	<i>Nerium indicum</i>	Apocyanaceae	Erlisch ascites carcinoma	5
39.	<i>Kallonji</i>	<i>Nigella sativa</i>	Ranunculaceae	Lewis lung carcinoma, Colon cancer	5
				Breast cancer cells	29
40.	<i>Tulsi</i>	<i>Ocimum sanctum</i>	Labiatae	Skin and liver tumors	29
				Dalton Lymphoma ascites (DLA) tumor model in Swiss Albino mice	35
41.	<i>Shyonak</i>	<i>Oroxylum indicum</i>	Bignoniaceae	HeLa cells	48
42.	<i>Gandhaprasa rini</i>	<i>Paederia foetida</i>	Rubiaceae	Human epidermoid carcinoma of the Nasopharynx	5
43.	<i>Katuki</i>	<i>Picrorrhiza kurroa</i>	Schrophulariaceae	Hepatic cancers	5
44.	<i>Bhumi amalaki</i>	<i>Phyllanthus niruri</i>	Phyllanthaceae	Skin melanoma and prostate cancer cells	49
45.	<i>Kankola</i>	<i>Piper cubeba</i>	Piperaceae	Breast cancer cell lines (MCF-7, MDA-MB-231, MDA-MB-468)	50
46.	<i>Pippali</i>	<i>Piper longum</i>	Piperaceae	DU-145 prostate, A549 lung, THP-1 leukemia, IGR-OVI-1 ovary and MCF-7 breast cancer cell line	51
47.	<i>Chitrak</i>	<i>Plumbago zeylanica</i>	Plumbaginaceae	Hepatoma	5
48.	<i>Ashoka</i>	<i>Polyalthia longifolia</i>	Annonaceae	Ehrlich's ascites tumor and Dalton's solid tumor, HeLa and MCF-7 cells	52
				HL-60 cells	53
49.	<i>Dadima</i>	<i>Punica granatum</i>	Punicaceae	Prostate cancer cell line	54
				Human breast cancer	55
				Mammary organ culture (mouse)	56
50.	<i>Mulaka</i>	<i>Raphanus sativum</i>	Cruciferae	MDA-MB-231 human breast cancer cells	57
51.	<i>Revandachini</i>	<i>Rheum emodi</i>	Polygonaceae	Breast cancer cells (MDA-MB-231)	58
				MDA-MB-435S(human breast carcinoma), Hep3B (human hepatocellular carcinoma) and PC-3 (human prostate cancer)	59
				MDA-MB-435S and Hep3B cell lines	60
52.	<i>Eranda</i>	<i>Ricinus communis</i>	Euphorbiaceae	SKMEL28 human melanoma	61

				cells	
53.	<i>Manjishtha</i>	<i>Rubia cordifolia</i>	Rubiaceae	P-388, L-1210, B-16 melanoma, colon 388, Lewis lung carcinoma, mammary carcinoma	5
54.	<i>Bhallataka</i>	<i>Semecarpus anacardium</i>	Anacardiaceae	Leukemia, melanoma and glioma, hepatocarcinoma	62
55.	<i>Kakamachi</i>	<i>Solanum nigrum</i>	Solanaceae	Human Leukemic Cell Lines	63
				HepG2 and CT26 cancer cell lines	64
				Human endometrial carcinoma cell lines, HEC1A, HEC1B, and KLE	65
56.	<i>Kantakari</i>	<i>Solanum xanthocarpum</i>	Solanaceae	leukemia (THP1) and lung cancer (HOP62) cell lines	66
57.	<i>Lavanga</i>	<i>Syzygium aromaticum</i>	Myrtaceae	Colon, Lung, and Breast Cancer Cell Lines	67
				MCF7 human breast cancer cell lines	68
				MCF-7 human estrogen dependent breast cancer and MDA-MB-231 human estrogen independent breast cancer cell lines (ATCC)	69
				Lung cancer	70
58.	<i>Jambu</i>	<i>Syzygium cumini</i>	Myrtaceae	Cervical cancer cell lines	71
				Skin papillomagenesis (mice)	72
				Gastric carcinogenesis(mice)	73
59.	<i>Talisapatra</i>	<i>Taxus baccata</i>	Taxaceae	Cytotoxic against various tumors	5
				HepG2 (Human hepatocarcinoma) and SKOV3 (Human ovary carcinoma)	74
				Human gastric and colon tissue	75
60.	<i>Sarpunkha</i>	<i>Tephrosia purpurea</i>	Fabaceae	MCF-7 cell line	29
				Human breast cancer cell line	41
61.	<i>Arjuna</i>	<i>Terminalia arjuna</i>	Combretaceae	Hepatocellular carcinoma	76
62.	<i>Bibhitaki</i>	<i>Terminalia bellerica</i>	Combretaceae	human cancer cell lines, including cancers of lung (A549), prostate (PC-3), breast (T47D and MCF-7), colon (HCT-16 and Colo-205) and leukemia (THP-1, HL-60 and K562)	77
63.	<i>Haritaki</i>	<i>Terminalia chebula</i>	Combretaceae	Lung Cancer Cells (A549) and mouse lung cancer cell line LLC	78
64.	<i>Guduchi</i>	<i>Tinospora cordifolia</i>	Menispermaceae	Dalton Lymphoma ascites (DLA) tumor model in Swiss Albino mice	35
65.	<i>Gokshur</i>	<i>Tribulus terrestris</i>	Zygophyllaceae	Liver cancer cells	79
				Papillomagenesis, HepG2 cells, human skin fibroblasts	80
66.	<i>Methika</i>	<i>Trigonella foenum-graecum</i>	Fabaceae	T-cell lymphoma (TCP), B-cell lymphomas, Thyroid Papillary carcinoma (FRO) and breast cancer (MCF7)	81
				Hepatocellular Carcinoma Cell Line, HepG2	82

				Breast, pancreatic and prostate cancer cell lines	83
67.	<i>Sadapushpi</i>	<i>Vinca rosea</i>	Apocyanaceae	P-1534, carcinoma of the breast, cervix, kidney, lung and ovary	5
68.	<i>Nirgundi</i>	<i>Vitex negundo</i>	Verbenaceae	Human hepatoma cells	84
				Brine shrimp lethality bioassay	85
69.	<i>Draksha</i>	<i>Vitis vinifera</i>	Vitaceae	Skin Cancer Cell Lines A431	86
				Oral squamous cell carcinoma	87
70.	<i>Ashwagandha</i>	<i>Withania somnifera</i>	Solanaceae	Lung adenomas(Mouse)	88
				Prostate(PC3), ovary(A2780), breast(MCF7), oral(DWD), lungs(Hop62) and colon(Colo205)	89
71.	<i>Sunthi</i>	<i>Zingiber officinalis</i>	Zingiberaceae	B164A5 murine melanoma cell line	90
				Human colorectal cancer cells, human breast cancer cell lines (MCF-7 and MDA-MB231) and human hepatocellular carcinoma (HepG-2)	91
				Liver cancer(Rats)	92
				Human breast carcinoma cell lines	93
72.	<i>Badar</i>	<i>Ziziphus mauritiana</i>	Rhamnaceae	Dalton Lymphoma ascites (DLA) tumor model in Swiss Albino mice	35

Phytochemical classes and cancer

Alkaloids

Alkaloids are one of the major classes of compounds possessing anticancer activity. Several alkaloids isolated from natural herbs exhibit antiproliferation and antimetastasis effects on various types of cancers both *in vitro* and *in vivo*.⁹⁴ One of the oldest and most important anticancer drugs found from the plant *Vinca rosea* i.e. *Vinca* alkaloids. Some other alkaloids colchicine, morphine, berberine etc found active against cancer.

Glycosides

In the 1960s inhibition of malignant cells of cardiac glycosides *in vitro* was reported and since then other anticancer effects of cardiac glycosides have been observed. Therapeutic effect of cardiac glycosides in breast cancer has been known from 1979.⁹⁵

Flavonoids

A huge number of epidemiological studies have been conducted to prove the protective effect of flavonoids against cancer. Flavonoids greatly influence the cascade of immunological events associated with the development and progression of cancer. *In vivo* studies using animal models have suggested the protective effect of flavonoids against initiation as well as tumor progression.⁹⁶

Phenolics

The antiproliferative effects (through apoptosis) induced by the phenolic agents on several cancer cell lines, for instance, were explained in terms of topoisomerase or phosphatidylinositol-3-kinase inhibition, or even cell cycle arrest. It is generally appreciated that the toxicity associated with some phenolic compounds is mediated by their oxidative activity, which can accelerate oxidative damage *in vitro*, either to DNA or to proteins and carbohydrates. Another possible, although insufficiently investigated, mechanism of phenol cytotoxicity may be associated with their pro-oxidant properties.⁹⁷

Saponins

Modern research found that saponins have antitumor effect on many cancer cells. Several saponins inhibit tumor cell growth by cell cycle arrest and apoptosis with IC₅₀ values upto 0.2 mM.⁹⁸

Essential oils

A very promising field of treatment with essential oils is their application against tumors. Especially since the 1990s the anticancer properties of essential oils and/or their main constituents and/or metabolites have gained more and more interest, in as much as such a “natural” therapy is accepted all over the world by the patients.⁹⁹

Terpenes

Terpenoids constitute the largest class of natural products and are a rich reservoir of candidate compounds for drug discovery. Recent efforts into the research and development of anti-cancer drugs derived from natural products have led to the identification of a variety of terpenoids that inhibit cancer cell proliferation and metastasis via various mechanisms.¹⁰⁰

Steroids

Phytosterols possess anti-cancer effects against cancer of the lung, stomach, ovary and estrogen-dependent human breast cancer. It has been speculated that phytosterols inhibit the production of carcinogens, cancer-cell growth, invasion and metastasis, and promote apoptosis of cancerous cells.¹⁰¹

Table 2 presents list of few different active plant constituents with their phytochemical class and anticancer activity which give an outline about main fundamental phytochemical classes which have potent anticancer activity.

Table 2. Active constituents with their anticancer activity

Active constituents	Plant source	Class	Activity	References
Berberine	<i>Berberis aristata</i>	Alkaloid	Human breast cancer T47D and MCF7 cell lines	102
			HONE1 cells (a human nasopharyngeal carcinoma cell line)	103
Colchicine	<i>Gloriosa superba</i>	Alkaloid	Leukemic and solid tumors	104
Flavopiridol	<i>Amoora rohituka</i>	Alkaloid	Head and neck squamous cell carcinomas	105
			Bladder Cancer Cells	106
Morphine	<i>Papaver somniferum</i>	Alkaloid	Human lung and breast carcinoma cell lines	107
Piperine	<i>Piper longum</i>	Alkaloid	Lung cancer	108
			Human fibrosarcoma (HT1080) cell, human breast cancer with <i>HER2</i> , human rectal tumor (HRT)18 cells, human prostate cancer cells	109
Pippalartine	<i>Piper longum</i>	Alkaloid	Prostate cancer cells, colon cancer cell	109
Taxol	<i>Taxus baccata</i>	Alkaloid	HeLa Cervical carcinoma, A549 Lung adenocarcinoma, U373 Grade III astrocytoma, MCF-7 Breast adenocarcinoma, HT-29 Colon adenocarcinoma, OVG-1 Ovarian carcinoma, PC-Sh Pancreatic adenocarcinoma, PC-Zr Pancreatic adenocarcinoma	110
Vincristine, Vinblastine	<i>Catharanthus rosea</i>	Alkaloid	Urothelial tract, nonsmall cell lung cancer and carcinoma of the breast	111

Bullatacin	<i>Annona muricata</i>	Glycoside	Hepatocarcinoma	112
			Mammary adenocarcinoma	113
Curcumin	<i>Curcuma longa</i>	Phenolics	Colon Carcinogenesis	114
			Mammary tumor	115
Gingerol	<i>Zingiber officinalis</i>		cancer cell lines including prostate, gastric, and breast	116
Resveratol	<i>Vitis vinifera</i>	Polyphenolics	Prostate cancer cell lines	117
			Human mammary epithelial cells	118
Diosgenin	<i>Trigonella foenum graecum</i>	Saponin	Rat colon carcinogenesis, HT-29 human colon cancer cell line	119
			Gastric Cancer BGC-823 Cells	120
Terrestrosin D	<i>Tribulus terrestris</i>	Saponin	Human prostate cancer	121
Anethole	<i>Foeniculum vulgare</i>	Essential oil	Human skin fibroblasts	122
Eugenol	<i>Syzygium aromaticum</i>	Essential oil	HL-60 human promyelocytic leukemia Cells	123
Thymoquinone	<i>Nigella sativa</i>	Essential oil	Colorectal carcinoma, breast adenocarcinoma, osteosarcoma, ovarian carcinoma, myeloblastic leukemia, and pancreatic carcinoma	116
Chebulagic acid	<i>Terminalia chebula</i>	Phenolic	Retinoblastoma cells	124
Gallic acid	<i>Terminalia bellerica, Emblica officinalis</i>	Phenolic	A549, a human lung adenocarcinoma cell line	125
			Gastric cancer cells	126
Urushiol	<i>Semecarpus anacardium</i>	Oleoresin	Breast cancer cells, MCF-7 and MDA-MB-231 cell lines	127
Emodin	<i>Rheum emodi</i>	Anthraquinone	HepG2, MDAMB231 and NIH/3T3 cells lines	128
Aloe-emodin	<i>Aloe vera</i>	Anthraquinone	CH27 (human lung squamous carcinoma cell) and H460 (human lung non-small cell carcinoma cell)	129
			Prostate cancer	130
			Breast cancer MCF-7 cells	131
			Human colon carcinoma cells	132
Boswellic acid	<i>Boswellia serrata</i>	Resin	Human malignant glioma cells	133
			Colon cancer cells	134
			Human prostate cancer xenografts in mice	135
			Colorectal cancer	136
Gambogic acid	<i>Garcinia indica</i>	Resin	Human hepatoma, breast cancer, gastric carcinoma, and lung carcinoma	116
Podophyllotoxin	<i>Podophyllum hexandrum</i>	Resin	ovarian, renal and lung cancer cell lines	137

Abrin	<i>Abrus precatorius</i>	Protein	B-cell lymphocytic-leukaemia	138
			Human small cell lung cancer	139
			Dalton's lymphoma ascites model	140
			MCF-7 (breast carcinoma), HepG2(hepatocarcinoma), KB (nasopharyngeal carcinoma)	141
			colon cancer LoVo cell in vitro	142
Allicin	<i>Allium sativum</i>	Protein	Human gastric cancer cell lines	143
			Human gastric cancer cell line SGC-7901	144
			Human ovarian cancer cell line	145
			Renal clear cell carcinoma	146
			Malignant Human B-Cells	147
Ricin	<i>Ricinus communis</i>	Protein	Human cervical cancer cells	148

Conclusion

Plants are good source of natural chemicals which are fruitful to fight against cancer. This paper reviews anticancer activity of different plants and group of phytochemicals representing good candidates for anticancer action. All compounds reviewed in the present work have a well documented anticancer activity in different cell lines. Phytochemicals possess multiple mechanisms to fight against cancer. By utilizing this compound as a part of the treatment survival rate of patient can be expanded. The main aim of the review fulfilled by the conclusion that the main phytochemical groups fighting against cancer with more number of candidates are alkaloids, phenolics and flavonoids. More research work can be performed towards these classes of phytochemicals and there is also scope to find out main active compound from the other plants which gave anticancer activity in crude extract form.

REFERENCES

- [1] Desai, A. G. et al., Medicinal Plants and Cancer Chemoprevention. *Curr Drug Metab.* 2008, 9, 581–591.
- [2] Jena, J., Ranjan, R., Ranjan, P., Sarangi, M. K., A Study On Natural Anticancer Plants. *International Journal Of Pharmaceutical And Chemical Sciences.* 2012, 1, 365-368.
- [3] Jain, R., Kosta, S., Tiwari, A., Ayurveda and cancer. *Pharmacognosy Res.* 2010, 2, 393–394.
- [4] Balachandran, P., Govindarajan, R., Cancer—an ayurvedic perspective. *Pharmacological Research*, 2005, 51, 19–30.
- [5] Dhiman, A., Hiremath, S. K., Pathak, M., Mishra, D., A review article on anticancerous drugs in ayurveda and screened anticancer activity of medicinal plants, *international journal of ayurveda & alternative medicine.* 2014, 2, 54-60.
- [6] Mohammed, S. S. et al., Cytotoxic and pro-apoptotic effects of *Abrus precatorius* L. on human metastatic breast cancer cell line, MDA-MB-231. *Cytotechnology*, 2013, 65, 407–417.
- [7] Jagetia, G. C., Venkatesh P., Baliga M. S., *Aegle marmelos* (L.) CORREA Inhibits the Proliferation of Transplanted Ehrlich Ascites Carcinoma in Mice. *Biol. Pharm. Bull.*, 2005, 28, 58–64.
- [8] Maity, P., Hansda, D., Bandyopadhyay, U., Mishra, D. K., Biological activities of crude extracts and chemical constituents of *Bael*, *Aegle marmelos*(L.) *Corr.* *Indian journal of Experimental Biology*, 2009, 47, 849-861.
- [9] Bhatti, R., Singh, J., Saxena, A. K., Suri, N., Ishar, M. P. S., Pharmacognostic standardisation and antiproliferative activity of *Aegle marmelos* (L.) *Correa* leaves in various human cancer cell lines. *Indian Journal of Pharmaceutical Sciences*, 2013, 75, 628–634
- [10] Varadharajan, S., Kalathil, K., Kuppusamy, A. K., Induction of Apoptosis and Cytotoxic Activities of *Apium graveolens* Linn. Using in vitro Models. *Middle-East Journal of Scientific Research*, 2011, 9, 90-94.
- [11] Muangnoi, P. et al., Cytotoxicity, Apoptosis and DNA Damage Induced by *Alpinia galanga* Rhizome Extract. *Planta medica*, 2007, 73, 748-754.
- [12] Wang, Y., Tian, W., Ma, X., Inhibitory Effects of Onion (*Allium cepa* L.) Extract on Proliferation of Cancer Cells and Adipocytes via Inhibiting Fatty Acid Synthase. *Asian Pacific J Cancer Prev*, 2012, 13, 5573-5579.
- [13] Jagetia, G. C., Baliga, M. S., Venkatesh, P., Effect of *Sapthaparna* (*Alstonia scholaris* Linn) in modulating the benzo(a) pyrene-induced forestomach carcinogenesis in mice, *Toxicology Letters*, 2003, 144, 183-193.
- [14] Jahan, S., Chaudhary, R., Goyal, P. K., Anticancer Activity of an Indian Medicinal Plant, *Alstonia scholaris*, on Skin Carcinogenesis in Mice. *Integrative Cancer Therapies*, 2009, 8, 273–279.
- [15] Ansil, P. N., Wills, P. J., Varun, R., Latha, M. S., Cytotoxic and apoptotic activities of *Amorphophallus campanulatus* (Roxb.) Bl. Tuber extracts against human colon carcinoma cell line HCT-15 Saudi. *Journal of Biological Sciences*, 2014, 21, 524–531.
- [16] Rajeshkumar, et al., Anticancer activity of *Andrographis paniculata* Leaves extract against neuroblastoma (IMR-32) and human colon (HT-29) cancer cell line. *World Journal of Pharmacy and Pharmaceutical Sciences*, 2015, 4, 1667-1675.
- [17] Suhesti, E. P. N. R. T. S., Adityono, R. W., The breast of anticancer from leaf extract of *Annona muricata* against cell line in T47D. *International Journal of Applied Science and Technology*, 2012, 2, 157-164.

- [18] Rosdi M. et al., Cytotoxic effect of *Annona muricata* Linn leaves extract on Capan-1 Cells. *Journal of Applied Pharmaceutical Science*, 2015, 5, 045-048.
- [19] Hamizah, S., Roslida, Ah., Fezah, O., Tan, KL., Tor, YS., Tan, CI., Chemopreventive Potential of *Annona muricata* L Leaves on Chemically-Induced Skin Papillomagenesis in Mice, *Asian Pacific Journal of Cancer Prevention*, 2012, 13, 2533-2539.
- [20] Suyatmi, Suselo, Y., H., Jusuf, S., A., The Selective Cytotoxicity Of Ethanolic Extract Of *Annona muricata* Leaf On HeLa Cervical Cancer Cells. *International Conference: Research and Application on Traditional Complementary and Alternative Medicine in Health Care (TCAM)*, 2012, 24-27.
- [21] Gavamukulya, Y., Abou-Elella, F., Wamunyokoli, F., AEl-Shemy, H., Phytochemical screening, anti-oxidant activity and in vitro anticancer potential of ethanolic and water leaves extracts of *Annona muricata* (Graviola). *Asian Pacific Journal of Tropical Medicine*, 2014, 7, S355-S363.
- [22] Verma, S. P., Tripathi, V. C., Das, P, *Asparagus racemosus* leaf extract inhibits growth of UOK 146 renal cell carcinoma cell line: simultaneous oncogenic PRCCTFE3 fusion transcript inhibition and apoptosis independent cell death. *Asian Pacific Journal of Cancer Prevention*, 2014, 15, 1937-1941.
- [23] Chaudhari, P., Vidyath, R., A Critical Appraisal of Herbs Useful in the Management of Cancer. *Int. J. Pharm. Phytopharmacol. Res.*, 2013, 2, 350-356.
- [24] Mallick, M. N. et al., Evaluation of anticancer potential of *Bacopa monnieri* L. against MCF7 and MDAMB 231 cell line, *J Pharm Bioallied Sci.* 2015, 7, 325-328.
- [25] Kilari, B. P., Kotakadi, V., S., Penchalaneni, J., Anti-proliferative and Apoptotic Effects of *Basella rubra* (L.) Against 1, 2-Dimethyl Hydrazine-induced Colon Carcinogenesis in Rats. *Asian Pacific Journal of Cancer Prevention*, 2016, 17, 73-80.
- [26] Kumar, S. S. et al., Fruit extracts of *Basella rubra* that are rich in bioactives and betalains exhibit antioxidant activity and cytotoxicity against human cervical carcinoma cells. *Journal of Functional Foods*, 2015, 15, 509-515.
- [27] Mishra, A. et al., *Bauhinia variegata* Leaf Extracts Exhibit Considerable Antibacterial, Antioxidant, and Anticancer Activities. *BioMed Research International*, 2013, 2013, 10 pages.
- [28] Morales, M. A. et al., Is Senna Laxative Use Associated to Cathartic Morales, M. A. et al., Is Senna Laxative Use Associated to Cathartic colon, Genotoxicity, or Carcinogenicity? *Journal of Toxicology*. 2009, 2009(287247), 1-8.
- [29] Singh, R. K., Singh, A. K., Atri, N., Singh, S. K., Traditionally used Medicinal Plants as Alternative Source for Future Anticancer Drugs. *The International Journal Of Science & Technoledge*, 2015, 3, 111-115.
- [30] Entezari, M. et al., Antimutagenicity and Anticancer Effects of Citrus Medica Fruit Juice. *Acta Medica Iranica*, 2009, 47, 373-377.
- [31] Bhandari, P. R., *Crocus sativus* L. (saffron) for cancer chemoprevention: A mini review. *J Tradit Complement Med.*, 2015, 5, 81-87.
- [32] Danciu, C. et al., Evaluation of phenolic profile, antioxidant and anticancer potential of two main representants of Zingiberaceae family against B164A5 murine melanoma cells. *Biological Research*, 2015, 48, 1.
- [33] Adhvaryu, M. R., Reddy, N., Parabia, M., Anti-Tumor Activity of Four Ayurvedic Herbs in Dalton Lymphoma Ascites Bearing Mice and Their Short-Term In Vitro Cytotoxicity on DLA-Cell-Line. *Afr J Tradit Complement Altern Med.*, 2008, 5, 409-418.
- [34] Kilani, S. et al., In vitro evaluation of antibacterial, antioxidant, cytotoxic and apoptotic activities of the tubers infusion and extracts of *Cyperus rotundus*. *Bioresour Technol.* 2008, 99, 9004-8.
- [35] Soumaya, K. J., Evaluation of in vitro antioxidant and apoptotic activities of *Cyperus rotundus*. *Asian Pac J Trop Med.* 2014, 7, 105-12.
- [36] Kilani-Jaziri, S. et al., Relationship correlation of antioxidant and antiproliferative capacity of *Cyperus rotundus* products towards K562 erythroleukemia cells. *Chem Biol Interact.* 2009, 181, 85-94.
- [37] Sancheti, G., Jindal, A., Kumari, R., Goyal, P. K., Chemopreventive action of *Emblca officinalis* on skin carcinogenesis in mice. *Asian Pacific Journal of Cancer Prevention*, 2005, 6, 197-201.
- [38] Zhao, T., Sun, Q., Marques, M., Witcher, M., Anticancer Properties of *Phyllanthus emblica* (Indian Gooseberry). *Hindawi Publishing Corporation Oxidative Medicine and Cellular Longevity* 2015, 2015, Article ID 950890, 7 pages.
- [39] De, A. et al., *Emblca officinalis* Extract Induces Autophagy and Inhibits Human Ovarian Cancer Cell Proliferation, Angiogenesis, Growth of Mouse Xenograft Tumors. *PLoS ONE* 2013, 8.
- [40] Khan, N., Sultana, S., Chemomodulatory effect of *Ficus racemosa* extract against chemically induced renal carcinogenesis and oxidative damage response in Wistar rats. *Life sciences*, 2005, 77, 1194-1210.
- [41] Gulecha, V., Sivakuma, T., Anticancer activity of *Tephrosia purpurea* and *Ficus religiosa* using MCF 7 cell lines. *Asian Pacific Journal of Tropical Medicine*, 2011, 2011, 526-529.
- [42] Manosroi, A. et al., Potent Antiproliferative Effect on Liver Cancer of Medicinal Plants Selected from the Thai/Lanna Medicinal Plant Recipe Database "MANOSROI III". *Evidence-Based Complementary and Alternative Medicine*, 2015, 2015, Article ID 397181, 11 pages.
- [43] Sharma, V., Hussain, S., Gupta, M., Saxena, A. K., In Vitro anticancer activity of extracts of *Mentha* Spp. Against human cancer cells. *Indian Journal of Biochemistry & Biophysics*, 2014, 51, 416-419.
- [44] Khan, M. et al., Hexane soluble extract of *Mallotus philippensis* (Lam.) Muell. Arg. root possesses anti-leukaemic activity. *Chemistry Central Journal*, 2013, 7.
- [45] Abdullah et al., Cytotoxic effects of *Mangifera indica* L. kernel extract on human breast cancer (MCF-7 and MDA-MB-231 cell lines) and bioactive constituents in the crude extract, *BMC Complementary and Alternative Medicine* 2014, 14, 199.
- [46] Garrido, G. et al., Polyphenols of *Mangifera indica* modulate arsenite-induced cytotoxicity in a human proximal tubule cell line. *Brazilian Journal of Pharmacognosy*, 2012, 22, 325-334.
- [47] Sharma, V., Hussain, S., Gupta, M., Saxena, A. K., In Vitro anticancer activity of extracts of *Mentha* Spp. Against human cancer cells. *Indian Journal of Biochemistry & Biophysics*, 2014, 51, 416-419.
- [48] Moirangthem, D. S. et al., Differential effects of *Oroxylum indicum* bark extracts: antioxidant, antimicrobial, cytotoxic and apoptotic study. *Cytotechnology*, 2013, 65, 83-95.
- [49] Tang, Y. Q., Jaganath, I. B., Sekaran, S. D., *Phyllanthus* spp. Induces Selective Growth Inhibition of PC-3 and MeWo Human Cancer Cells through Modulation of Cell Cycle and Induction of Apoptosis. 2010, *PLoS ONE*, 5, e12644.
- [50] Graidist, P., Martla, M., Sukpondma, Y., Cytotoxic Activity of *Piper cubeba* Extract in Breast Cancer Cell Lines. *Nutrients* 2015, 7, 2707-2718.
- [51] Sharma, A. K. et al., Cell cycle inhibitory activity of *Piper longum* against A549 cell line and its protective effect against metal-induced toxicity in rats. *Indian Journal of Biochemistry & Biophysics*, 2014, 51, 358-364.
- [52] Manjula, S. N. et al., Antitumor and antioxidant activity of *Polyalthia longifolia* stem bark ethanol extract. *Pharmaceutical Biology*, 2010, 48, 690-696.
- [53] Verma, M. et al., In vitro cytotoxic potential of *Polyalthia longifolia* on human cancer cell lines and induction of apoptosis through mitochondrial-dependent pathway in HL-60 cells. *Chemo-biological Interactions*, 2008, 171, 45-56.

- [54] Albrecht, M. et al., Pomegranate Extracts Potently Suppress Proliferation, Xenograft Growth, and Invasion of Human Prostate Cancer Cells. *J Med Food* 2004, 7, 274-283.
- [55] Kim, N. D. et al., Chemopreventive and adjuvant therapeutic potential of pomegranate (*Punica granatum*) for human breast cancer. *Breast Cancer Research and Treatment*, 2002, 71, 203-217.
- [56] Mehta, R., Lansky, E., P., Breast cancer chemopreventive properties of pomegranate (*Punica granatum*) fruit extracts in a mouse mammary organ culture. *European Journal of Cancer Prevention*, 2004, 13, 345-348.
- [57] Kim, W. K. et al., Radish (*Raphanus sativus* L. leaf) ethanol extract inhibits protein and mRNA expression of ErbB2 and ErbB3 in MDA-MB-231 human breast cancer cells. *Nutrition Research and Practice*, 2011, 5, 288-293.
- [58] Kumar, D. R. N., George, V. C., Suresh, P. K., Kumar, R. A., Acceleration of pro-caspase-3 maturation and cell migration inhibition in human breast cancer cells by phytoconstituents of rheum emodi rhizome extracts. *EXCLI Journal* 2013, 12, 462-478.
- [59] Rajkumar, V., Guha, G., Kumar, R. A., Apoptosis induction in MDA-MB-435S, Hep3B and PC-3 cell lines by Rheum emodi rhizome extracts. *Asian Pacific Journal of Cancer Prevention*, 2011, 12, 1197-1200.
- [60] Rajkumar, V., Guha, G., Kumar, R. A., Antioxidant and Anti-Cancer Potentials of Rheum emodi Rhizome Extracts. *Evidence-Based Complementary and Alternative Medicine*, 2011, 2011, Article ID 697986, 9 pages.
- [61] Darmanin, S. et al., An extract from *Ricinus communis* L. leaves possesses cytotoxic properties and induces apoptosis in SKMEL28 human melanoma cells. *Natural Product Research: Formerly Natural Product Letters*, 2009, 23, 561-571.
- [62] Chaudhari, P., Vidyantath, R., A Critical Appraisal of Herbs Useful in the Management of Cancer. *Int.J.Pharm.Phytopharmacol.Res.*, 2013, 2, 350-356.
- [63] Gabrani, R. et al., Antiproliferative Effect of *Solanum nigrum* on Human Leukemic Cell Lines. *Indian J Pharm Sci.* 2012, 74, 451-453.
- [64] Shokrzadeh, M. et al., Cytotoxicity of hydroalcoholic extracts of *Cucurbita pepo* and *Solanum nigrum* on HepG2 and CT26 cancer cell lines. *Pharmacogn Mag.* 2010, 6, 176-179.
- [65] Tai, C. J. et al., Aqueous Extract of *Solanum nigrum* Leaf Activates Autophagic Cell Death and Enhances Docetaxel-Induced Cytotoxicity in Human Endometrial Carcinoma Cells. *Evid Based Complement Alternat Med.*, 2012: 859185.
- [66] Kumar, S., Pandey, A. K., Medicinal attributes of *Solanum xanthocarpum* fruit consumed by several tribal communities as food: an in vitro antioxidant, anticancer and anti HIV perspective. *BMC Complement Altern Med.*, 2014, 14: 112.
- [67] Hasan, A. U., Cytotoxic Activity of Curcumin, Melissa, and Cloves Extracts on Colon, Lung, and Breast Cancer Cell Line. *ETD Collection for Tennessee State University*, 2015, Paper AAI1585791.
- [68] Kumar, P. S. et al., Anticancer potential of *Syzygium aromaticum* L. in MCF7 human breast cancer cell lines. *Pharmacognosy Res.*, 2014, 6, 350-354.
- [69] Abdalrahim, F. A. et al., *Syzygium aromaticum* extracts as good source of betulinic acid and potential anti-breast cancer. *Brazilian Journal of Pharmacognosy*, 2012, 22, 335-343.
- [70] Banerjee, S., Panda, C. K., Das, S., Clove (*Syzygium aromaticum* L.), a potential chemopreventive agent for lung cancer. *Carcinogenesis*, 2008, 27, 1645-1654.
- [71] Barh, D., Vishwanathan, G., *Syzygium cumini* inhibits growth and induces apoptosis in cervical cancer cell lines: a primary study. *ecancer* 2008, 2:83.
- [72] Parmar, J., Sharma, P., Verma, P., Goyal, P. K., Chemopreventive action of *Syzygium cumini* on DMBA-induced skin papillomagenesis in mice. *Asian Pacific Journal of Cancer Prevention*, 2010, 11, 261-265.
- [73] Goyal, P. K., Verma, P., Sharma, P., Parmar, J., Agarwal, A., Evaluation of anti-cancer and anti-oxidative potential of *Syzygium Cumini* against benzo[a]pyrene (BaP) induced gastric carcinogenesis in mice. *Asian Pacific Journal of Cancer Prevention*, 2010, 11, 753-758.
- [74] Shokrzadeh, M. et al., Comparison of the cytotoxic effects of *Juniperus sabina* and *Zataria multiflora* extracts with *Taxus baccata* extract and Cisplatin on normal and cancer cell lines. *Pharmacogn Mag.*, 2010, 6, 102-105.
- [75] Durak, Z. E. et al., Aqueous extract from *Taxus baccata* inhibits adenosine deaminase activity significantly in cancerous and noncancerous human gastric and colon tissues. *Pharmacogn Mag.* 2014, 10, S214-S216.
- [76] Sivalokanathan, S., Ilayaraja, M., Balasubramanian, Efficacy of *Terminalia arjuna*(Roxb.) on N-nitrosodiethylamine induced hepatocellular carcinoma in rats. *Indian Journal of Experimental Biology*, 2005, 43, 264-267.
- [77] Diab, K. A., Guru, S. K., Bhushan, S., Saxena, A. K., In Vitro Anticancer Activities of *Anogeissus latifolia*, *Terminalia bellerica*, *Acacia catechu* and *Moringa oleifera* Indian Plants. *Asian Pacific Journal of Cancer Prevention*, 2015, 16, 6423-6428.
- [78] Wang, M. et al., Aqueous Extract of *Terminalia chebula* Induces Apoptosis in Lung Cancer Cells Via a Mechanism Involving Mitochondria-mediated Pathways. *Braz. Arch. Biol. Technol.* 2015, 58, 208-215.
- [79] Kim, H. J. et al., Aqueous extract of *Tribulus terrestris* Linn induces cell growth arrest and apoptosis by downregulating NFκB signaling in liver cancer cells. *Journal of Ethnopharmacology*, 2011, 136, 197-203.
- [80] Chhatre, S., et al., Phytopharmacological overview of *Tribulus terrestris*. *Pharmacogn Rev.* 2014, 8, 45-51.
- [81] Alsemari, A. et al., The selective cytotoxic anti-cancer properties and proteomic analysis of *Trigonella Foenum-Graecum*. *BMC Complementary and Alternative Medicine* 2014, 14:114.
- [82] Khalil, M. I. M., Ibrahim, M. M., El-Gaaly, G. A., Sultan, A. S., *Trigonella foenum* (Fenugreek) Induced Apoptosis in Hepatocellular Carcinoma Cell Line, HepG2, Mediated by Upregulation of p53 and Proliferating Cell Nuclear Antigen. *BioMed Research International*, 2015, 2015, Article ID 914645, 11 pages.
- [83] Shabbeer, S. et al., Fenugreek: a naturally occurring edible spice as an anticancer agent. *Cancer Biol Ther.*, 2009, 8, 272-278.
- [84] Kadir, F. A., Kassim, N., M., Abdulla, M. A., Yehye, W. A., PASS-predicted *Vitex negundo* activity: antioxidant and antiproliferative properties on human hepatoma cells-an in vitro study. *BMC Complementary and Alternative Medicine*, 2013, 13:343.
- [85] Khan, M. S. S. et al., Screening and evaluation of antioxidant, antimicrobial, cytotoxic, thrombolytic and membrane stabilizing properties of the methanolic extract and solvent-solvent partitioning effect of *Vitex negundo* Bark. *Asian Pac J Trop Dis*, 2013, 3, 393-400.
- [86] Mohansrinivasan, V., Exploring the Anticancer Activity of Grape Seed Extract on Skin Cancer Cell Lines A431. *Braz. Arch. Biol. Technol.*, 2015, 58, 540-546.
- [87] Aghbali, A., Induction of apoptosis by grape seed extract (*Vitis vinifera*) in oral squamous cell carcinoma. *Bosn J Basic Med Sci.*, 2013, 13, 186-191.
- [88] Monteiro, L. D. S. et al., Medicinal Plants and Other Living Organisms with Antitumor Potential against Lung Cancer, Evidence-Based Complementary and Alternative Medicine, 2014, 2014, Article ID 604152, 15 pages.
- [89] Gaidhani, S. N. et al., Evaluation of some plant extracts for standardization and anticancer activity. *Indian Journal of Traditional Knowledge*, 2013, 12, 682-687.
- [90] Danciu, C. et al., Evaluation of phenolic profile, antioxidant and anticancer potential of two main representants of Zingiberaceae family against B164A5 murine melanoma cells. *Biological Research* 2015, 48, 1-9.

- [91] Park, G. H. et al., Anti-cancer activity of Ginger (*Zingiber officinale*) leaf through the expression of activating transcription factor 3 in human colorectal cancer cells. *BMC Complementary and Alternative Medicine* 2014, 14, 408.
- [92] Yusof, M. et al., Chemopreventive Efficacy Of Ginger (*Zingiber Officinale*) In Ethionine Induced Rat Hepatocarcinogenesis. *Afr. J. Trad. CAM* 2009, 6, 87 – 93.
- [93] Rahman, S., Salehin, F., Iqbal, A., In vitro antioxidant and anticancer activity of young *Zingiber officinale* against human breast carcinoma cell lines. *BMC Complement Altern Med.*, 2011, 11, 76
- [94] Lu, J. J. et al., Alkaloids Isolated from Natural Herbs as the Anticancer Agents. *Evidence-Based Complementary and Alternative Medicine*, 2012, 2012, Article ID 485042, 12 pages
- [95] Winnicka, K., Bielawski, K., Bielawska, A., Cardiac Glycosides In Cancer Research and Cancer Therapy. *Acta Poloniae Pharmaceutica n Drug Research*, 2006, 63, 109-115.
- [96] Batra, P., Sharma, A. K., Anti-cancer potential of flavonoids: recent trends and future perspectives. *Biotech*, 2013, 3, 439–459.
- [97] Gomes, C. A. et al., Anticancer Activity of Phenolic Acids of Natural or Synthetic Origin: A Structure-Activity Study. *J. Med. Chem.* 2003, 46, 5395-5401.
- [98] Man, S. et al., Chemical study and medical application of saponins as anti-cancer agents. *Fitoterapia*, 2010, 81, 703–714.
- [99] Baser, K. H. C., Buchbauer, G., *Handbook of essential oils science, technology, and applications*. CRC Press Taylor & Francis Group, 6000 Broken Sound Parkway NW, Suite 300, pg no. 236.
- [100] Huang, M. et al., Terpenoids: natural products for cancer therapy. *Expert Opinion on Investigational Drugs*, 2012, 21, 1801-1818.
- [101] Woyengo, T. A., Ramprasath, V. R., Jones, P. J. H., Anticancer effects of phytosterols. *European Journal of Clinical Nutrition*, 2009, 63, 813–820.
- [102] Barzegar, E. et al., Effects of berberine on proliferation, cell cycle distribution and apoptosis of human breast cancer T47D and MCF7 cell lines. *Iran J Basic Med Sci*, 2015, 18, 334-342.
- [103] Tsang, C. M. et al., Berberine inhibits Rho GTPases and cell migration at low doses but induces G2 arrest and apoptosis at high doses in human cancer cells. *International Journal Of Molecular Medicine*, 2009, 24, 131-138.
- [104] Dubey, K. K., Ray, A. R., Behera, B. K., Production of demethylated colchicines through microbial transformation and scale-up process development. *Process Biochem.*, 2008, 43, 251-257.
- [105] Patel, V. et al., Flavopiridol, a Novel Cyclin-dependent Kinase Inhibitor, Suppresses the Growth of Head and Neck Squamous Cell Carcinomas by Inducing Apoptosis, *The Journal of Clinical Investigation*, 1998, 102, 1674-1681.
- [106] Wirger, A. et al., Flavopiridol, an Inhibitor of Cyclin-dependent Kinases, Induces Growth Inhibition and Apoptosis in Bladder Cancer Cells In Vitro and In Vivo, *Anticancer Research*, 2005, 25, 4341-4348.
- [107] Hatsukari, I. et al., Induction of early apoptosis marker by morphine in human lung and breast carcinoma cell lines. *Anticancer Research*, 2003, 23, 2413-2417.
- [108] Selvendiran, K., et al., Cytoprotective effect of piperine against benzopyrene induced lung cancer with reference to lipid peroxidation and antioxidant system in Swiss albino mice. *Fitoterapia*.2003, 74, 109-15.
- [109] Wang, Y. H. et al., Anticancer Principles from Medicinal Piper (胡椒 Hú Jiāo) Plants, *J Tradit Complement Med.* 2014 , 4, 8–16.
- [110] Liebmann, J. E. et al., Cytotoxic studies of paclitaxel (Taxol®) in human tumour cell lines. *Br. J. Cancer*, 1993, 68, 1104-1109.
- [111] Moudi, M., Go, R., Yien, C. Y. S., Nazre, M., Vinca Alkaloids, *Int J Prev Med.*, 2013, 4, 1231–1235.
- [112] Chih, H. W. et al., Bullatacin, a potent antitumor annonaceous acetogenin, inhibits proliferation of human hepatocarcinoma cell line 2.2.15 by apoptosis induction. *Life Sciences*, 2001, 69, 1321–1331.
- [113] Oberlies, N. H. et al., The Annonaceous acetogenin bullatacin is cytotoxic against multidrug-resistant human mammary adenocarcinoma cells. *Cancer Letters* 1997, 115, 73-79.
- [114] Rao, C. S., Rivenson, A., Simi, B., Reddy, B. S., Chemoprevention of Colon Carcinogenesis by Dietary Curcumin, a Naturally Occurring Plant Phenolic Compound, *Cancer Research* 1955, 55, 259-266.
- [115] Inano H et al, Chemoprevention by curcumin during the promotion stage of tumorigenesis of mammary gland in rats irradiated with g-rays, *Carcinogenesis*, 20 , 1011–1018.
- [116] Aggarwal, B. B. et al., Potential of Spice-Derived Phytochemicals for Cancer Prevention. *Planta Med*, 2008, 74, 1560–1569.
- [117] Kuwajerwala, N.; Cifuentes, E.; Gautam, S.; Menon, M.; Barrack, E.R.; Reddy, G.P.V. Resveratrol induces prostate cancer cell entry into S phase and inhibits DNA synthesis. *Cancer Res.* 2002, 62, 2488–2492.
- [118] Subbaramaiah, K. et al., Resveratrol inhibits cyclooxygenase-2 transcription and activity in phorbol ester-treated human mammary epithelial cells. *J. Biol. Chem.* 1998, 273, 21875–21882.
- [119] Alsemari, A., The selective cytotoxic anti-cancer properties and proteomic analysis of *Trigonella Foenum-Graecum*. *BMC Complementary and Alternative Medicine*, 2014, 14:114.
- [120] Mao, Z. J. et al., Anti-Proliferation and Anti-Invasion Effects of Diosgenin on Gastric Cancer BGC-823 Cells with HIF-1 α shRNAs. *Int. J. Mol. Sci.* 2012, 13, 6521-6533.
- [121] Wei, S. et al., Terrestrosin D, a steroidal saponin from *Tribulus terrestris* L., inhibits growth and angiogenesis of human prostate cancer in vitro and in vivo. *Pathobiology: Journal of Immunopathology, Molecular and Cellular Biology*, 2014, 81, 123-132.
- [122] Galicka, A., Kre, towski, R., Nazaruk, J., Cechowska-Pasko, M., Anethole prevents hydrogen peroxide-induced apoptosis and collagen metabolism alterations in human skin fibroblasts, *Mol Cell Biochem*, 2014, 394, 217–224.
- [123] H. Carrasco, A. et al., Eugenol and its Synthetic Analogues Inhibit Cell Growth of Human Cancer Cells (Part I). *J. Braz. Chem. Soc.*, 2008, 19, 543-548.
- [124] Kumar, N., Gangappa, D., Gupta, G., Roy, K., Chebulagic acid from *Terminalia chebula* causes G1 arrest, inhibits NF κ B and induces apoptosis in retinoblastoma cells, *BMC Complementary and Alternative Medicine* 2014, 14:319.
- [125] Maurya, D. K., Nandakumar, N., Devasagayam, T. P. A., Anticancer property of gallic acid in A549, a human lung adenocarcinoma cell line, and possible mechanisms, *J. Clin. Biochem. Nutr.*, 2011, 48, 85–90.
- [126] Ho, H. H. et al., Gallic acid inhibits gastric cancer cells metastasis and invasive growth via increased expression of RhoB, downregulation of AKT/small GTPase signals and inhibition of NF κ B activity. *Toxicology and Applied Pharmacology*, 2013, 266, 76–85.
- [127] Zhao, W. et al., Identification of urushiols as the major active principle of the Siddha herbal medicine *Semecarpus Lehyam*: Antitumor agents for the treatment of breast cancer, *Pharmaceutical Biology*, 2009, 47, 886–893.
- [128] Narender, T. et al., Preparation of novel antiproliferative emodin derivatives and studies on their cell cycle arrest, caspase dependent apoptosis and DNA binding interaction. *Phytomedicine*, 2013, 20, 890–896.
- [129] Lee, H. Z., Protein kinase C involvement in aloe-emodin- and emodin-induced apoptosis in lung carcinoma cell. *British Journal of Pharmacology*, 2001, 134, 1093 -1103.
- [130] Liu, K. et al., Aloe-emodin suppresses prostate cancer by targeting the mTOR complex 2. *Carcinogenesis*, 33, 1406–1411.
- [131] Chena, Q. et al., Exploring a novel target treatment on breast cancer: aloe-emodin mediated photodynamic therapy induced cell apoptosis and inhibited cell metastasis. *Anti-cancer Agents in Medicinal Chemistry*, 2015.

- [132] Lin, K. Y., Uen, Y. H., Aloe-emodin, an anthraquinone, in vitro inhibits proliferation and induces apoptosis in human colon carcinoma cells. *Oncology Letters*, 2010, 1, 541-547.
- [133] Glaser, T. et al., Boswellic acids and malignant glioma: induction of apoptosis but no modulation of drug sensitivity. *British Journal of Cancer*, 1999, 80, 756-765.
- [134] Liu, J., Huang, B., Hooi, S. C., Acetyl-keto-b-boswellic acid inhibits cellular proliferation through a p21-dependent pathway in colon cancer cells. *British Journal of Pharmacology*, 2006, 148, 1099-1107.
- [135] Pathania, M. et al., The anti-angiogenic and cytotoxic effects of the boswellic acid analog BA145 are potentiated by autophagy inhibitors. *Molecular Cancer* 2015, 14:6.
- [136] Takahashi, M. et al., Boswellic acid exerts antitumor effects in colorectal cancer cells by modulating expression of the let-7 and miR-200 microRNA family. *Carcinogenesis*, 2012, 33, 2441-2449.
- [137] Giri, A., Narasu, M. L., Production of podophyllotoxin from *Podophyllum hexandrum*: a potential natural product for clinically useful anticancer drugs. *Cytotechnology*, 2000, 34, 17-26.
- [138] Narayanan, S., Surolia, A., Karande, A. A., Ribosome-inactivating protein and apoptosis: abrin causes cell death via mitochondrial pathway in Jurkat cells. *Biochem. J.*, 2004, 377, 233-240.
- [139] Wawrzynczak, E. J. et al., Molecular and biological properties of an abrin A chain immunotoxin designed for therapy of human small cell lung cancer. *Br. J. Cancer*, 1992, 66, 361-366.
- [140] Bhutia, S. K., Mallick, S. K., Maiti, S., Maiti, T., Inhibitory effect of *Abrus* abrin-derived peptide fraction against Dalton's lymphoma ascites model. *Phytomedicine*. 2009, 16, 377-385.
- [141] Gadadhar, S., Karande, A. A., *Abrin* Immunotoxin: Targeted Cytotoxicity and Intracellular Trafficking Pathway, 2013, *PLoS ONE*, 8, e58304. doi:10.1371/journal.pone.0058304.
- [142] Gao, Y. et al., Allicin enhances cytotoxicity of CPT-11 to colon cancer LoVo cell in vitro. *China Journal of Chinese Materia Medica*. 2009, 34, 3092-3095.
- [143] Ha, M. W., Yuan, Y., Allicin induced cell cycle arrest in human gastric cancer cell lines. *Chinese Journal of Oncology*, 2004, 26, 585-589.
- [144] Tao, M., Gao, L., Pan, J., Wang, X., Study on the inhibitory effect of allicin on human gastric cancer cell line SGC-7901 and its mechanism. *Afr J Tradit Complement Altern Med.*, 2014, 11, 176-179.
- [145] Xu, L. et al., Role of JNK Activation and Mitochondrial Bax Translocation in Allicin-Induced Apoptosis in Human Ovarian Cancer SKOV3 Cells. *Evidence-Based Complementary and Alternative Medicine*, 2014, Article ID 378684, 6 pages.
- [146] Song, B., Shu, Y., Cui, T., Fu, P., Allicin inhibits human renal clear cell carcinoma progression via suppressing HIF pathway. *Int J Clin Exp Med* 2015, 8, 20573-20580.
- [147] Ghetie, M. A. et al., Evaluation of Ricin A Chain-containing Immunotoxins Directed against CD19 and CD22 Antigens on Normal and Malignant Human B-Cells as Potential Reagents for in Vivo Therapy. *Cancer Research*, 1988, 48, 2610-2617.
- [148] Rao, P. V. L., Mechanism of ricin-induced apoptosis in human cervical cancer cells. *Biochemical Pharmacology*, 2005, 69, 855-865.